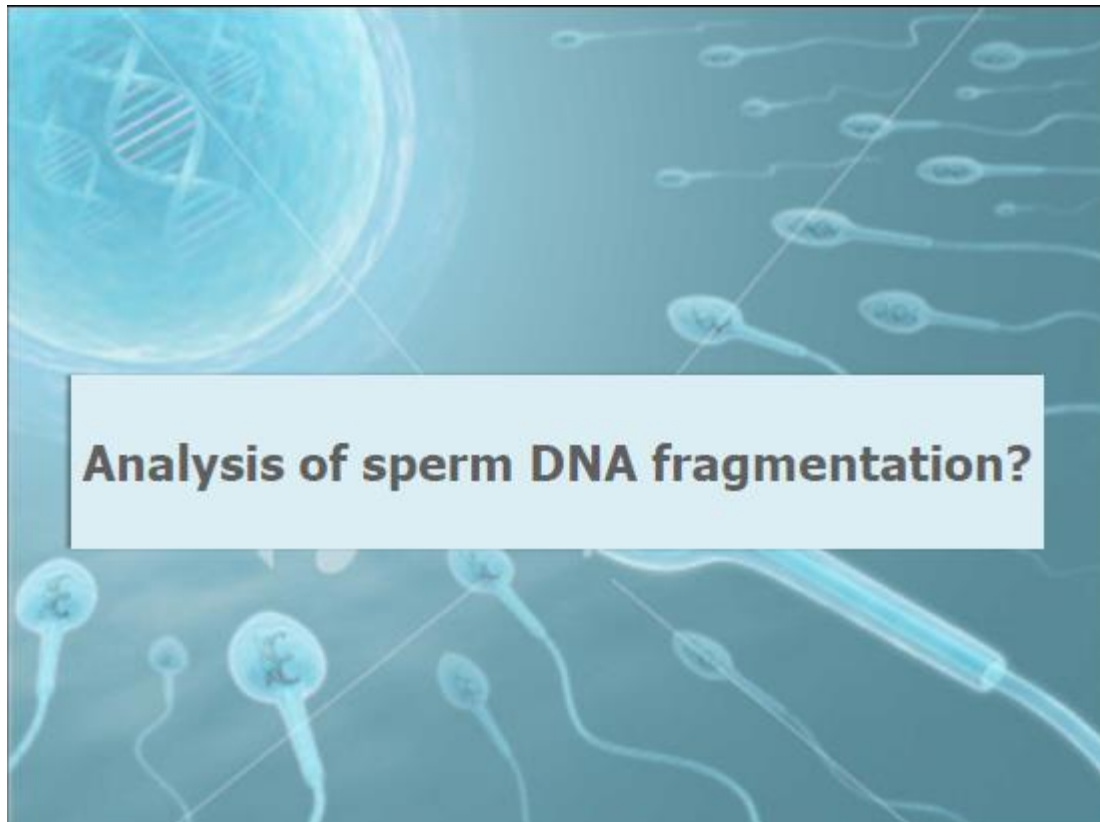


# 2nd Symposium of the Croatian Society of Clinical Embryologists and Andrology Workshop

Opatija, 29.12. 2014.

**COURSE:** **Sperm DNA fragmentation**

**MODERATORS:** doc dr sc Marjan Tandara (KBC Split),  
dr sc Sanja Vujisić (Poliklinika BetaPlus)  
mag biol mol Tea Rogić (Poliklinika BetaPlus)



## Male infertility etiology

- **Varicocele** (35-40%)
- **Infection** (~10%)
- **Genetics** (~10%)
- **Endocrinology** (<5%)
- **Immunology** (<5%)
- **Obstruction** (<5%)
- **Cryptorchidism** (<5%)
- **Idiopathic** (25%)

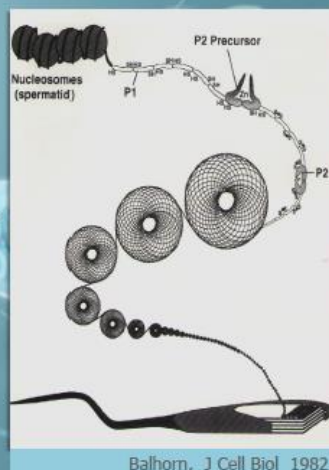
Greenberg et al, J Urology 1978

## Potential causes of DNA fragmentation

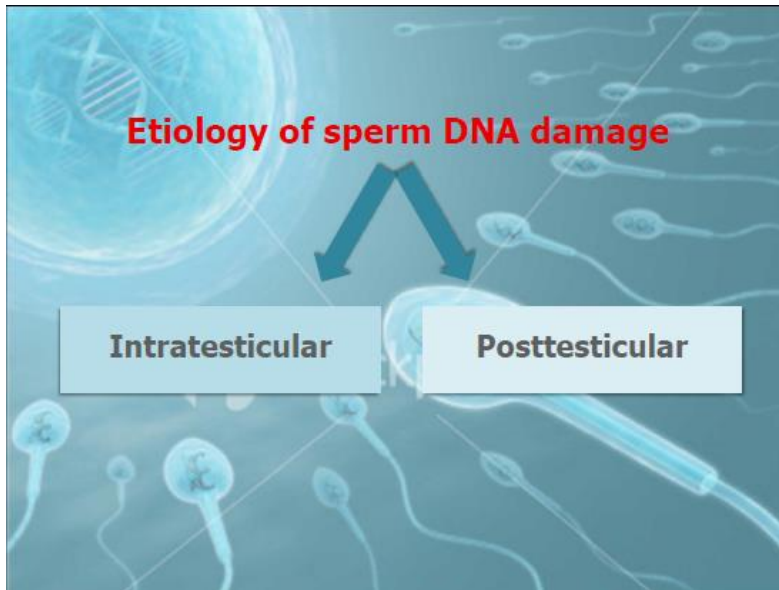
- **Varicocele** (Saleh, 2003; Fischer, 2003; Zini, 2000)
- **Chemotherapy and radiotherapy** (Chatterjee, 2000; Deane, 2004; Kobayashi, 2001)
- **Smoking** (Mak, 2000; Kunzle, 2003; Potts, 1999, Sun, 1993)
- **Leukocytospermia** (de Laminrande, 1993; Zini, 1995, Twigg, 1998; Iwasaki, 1992; Zini, 1993)
- **Apoptosis** (Baccetti, 1996; Sakkas, 2003)
- **Lack of protamine** (Cho, 2003)
- **Testicular hyperthermia** (Sailer J 1997; Banks Reproduction 2005)

## Sperm DNA packaging

- Binding of protamine (P1 i P2) on DNA replaces all other proteins (histones)
- Formation of ring structures (each consists of 50 Kbp DNA)
- Each spermatozoa has approximately 50 000 ring structures

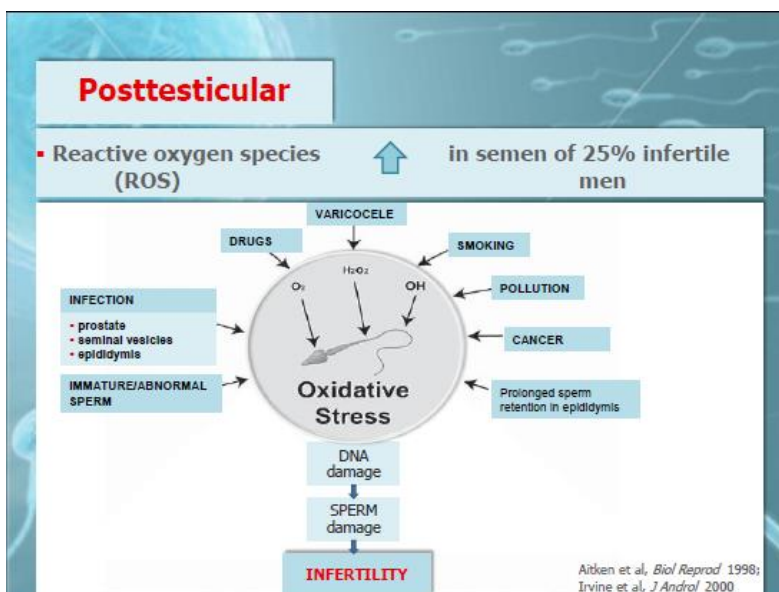


Balhorn, J Cell Biol 1982



### Intratesticular

- **Lack of protamine - a significant cause of sperm DNA damage (found in 5-15% of infertile men)**  
De Yebra et al, J Biol Chem 1993; Bianchi et al, Biol Reprod 1993; Kramer et al, Genet Test 1997, Cho, 2003
- **Mutations in genes coding for protamine synthesis**
- **Impaired topoisomerase II and transition proteins may cause DNA damage**  
Balhorn, J Cell Biol 1982; Kierzenbaum, Mol Reprod Dev 2001; Bissoneault, FEBS Lett 2002
- **Apoptosis**  
Sakkas et al, Exp Cell Res 1999, Sakkas et al. Reprod Biomed Online 2003





## Why investigate sperm DNA integrity?

- Need for better marker of male fertility potential
- Improvement of ART arised issues of sperm quality importance

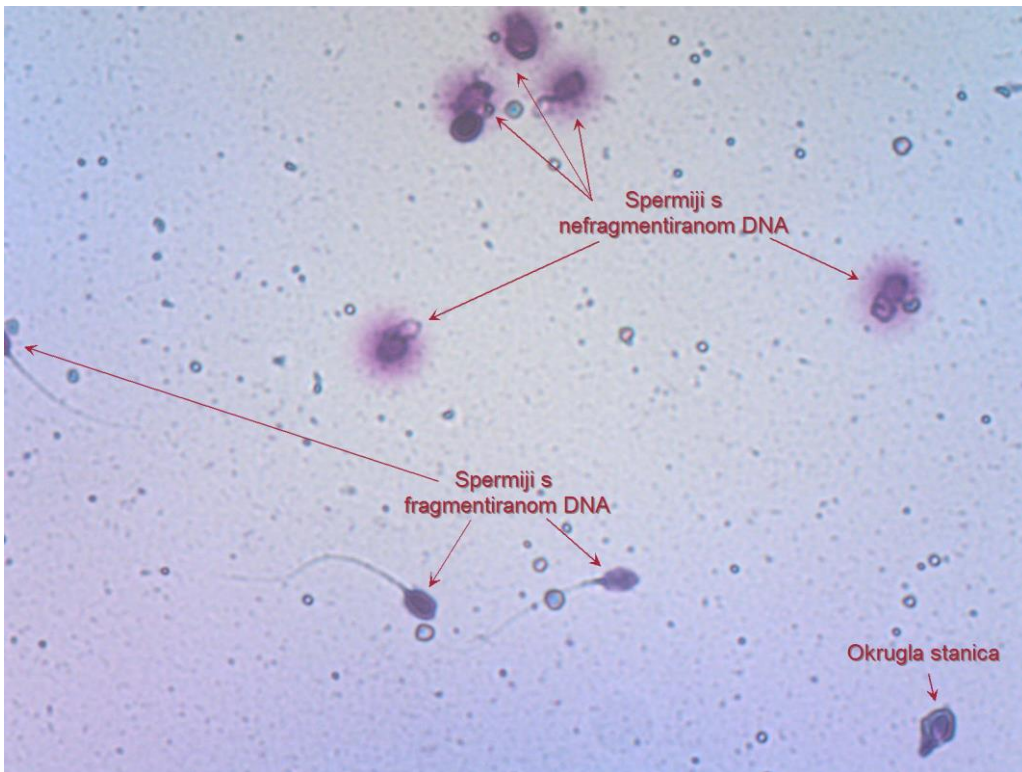
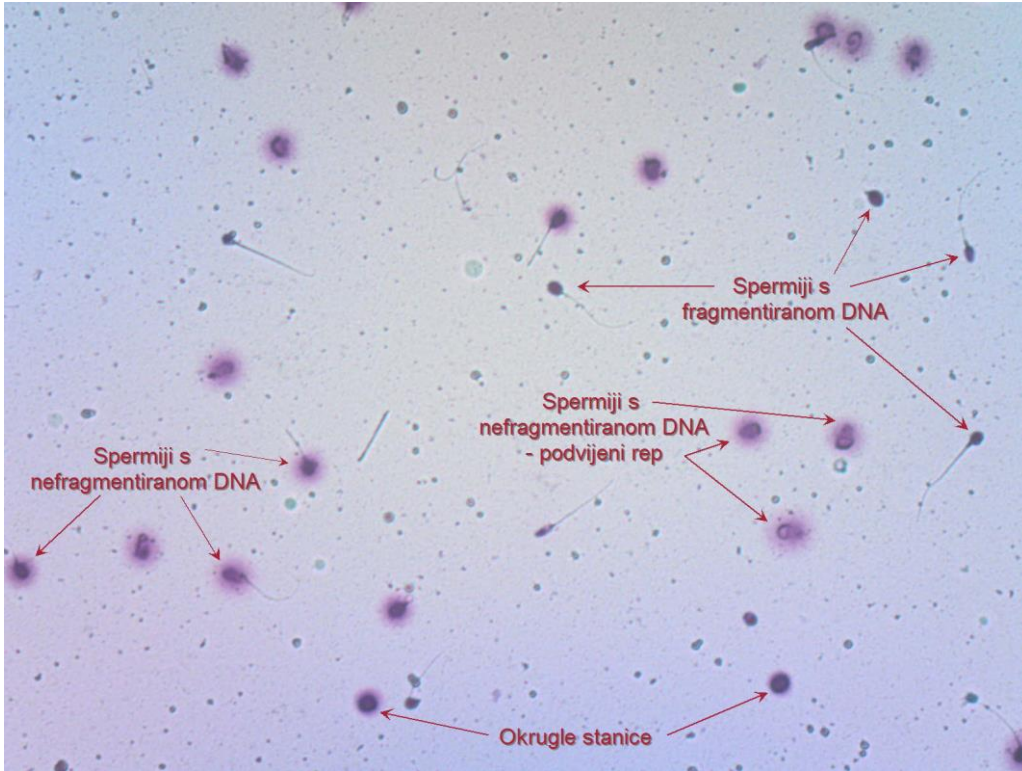


## DNA fragmentation analysis....

....is recommended for

- Patients with more than 3 consecutive unsuccessful IVF procedures
- In cases of embryo development arrest
- In cases of reccurent miscarriage
- In cases of idiopathic infertility





Alternative techniques for determining the integrity of sperm DNA (SCSA, TUNNEL, ISNT, comet assay) are quite labour-intensive and expensive, which prevents their inclusion in the standard spermiogram used in clinical praxis

## **PROCEDURE:**

Sperm DNA fragmentation can be determined using Halosperm® Sperm DNA Analysis Kit, which utilizes sperm chromatin dispersion (SCD) test (Fernández i sur., 2005) for detection of spermatozoa with poor quality DNA.

- 1) Prepare adequate sperm concentrations of  $5-10 \times 10^6/\text{mL}$  using sperm cultivation medium
- 2) Warm the Lysis Solution (LS) at the room temperature
- 3) Place the eppendorf tube with agarose (ACS) into the float and incubate in a water bath at  $95-100^\circ\text{C}$  for 5 min
- 4) Equilibrate the agarose in a water bath for another 5 min at  $+37^\circ\text{C}$
- 5) Add 25  $\mu\text{L}$  of sample in eppendorf tube with agarose and mix well. Place a drop of 20  $\mu\text{L}$  of sperm suspension on a processed side of the microscope slide (SCS) and cover with coverslip
- 6) Slides should be kept horizontally during the entire process
- 7) Place the slide on a surface pre-cooled to  $+4^\circ\text{C}$ , transfer into the fridge and leave for 5 min to allow for agarose to solidify
- 8) Prepare the denaturation solution by adding 80  $\mu\text{L}$  of Acid Denaturation (AD) solution to 10 mL of distilled water
- 9) Remove the coverslips from the slides and place the slides horizontally into the denaturation solution (in 100 mm Petri plates) and incubate 7 min
- 10) Transfer the slides into next Petri plate and incubate in 10 mL of Lysis solution for 25 min
- 11) Transfer the slides into next Petri plate and incubate in distilled water for 5 min
- 12) Incubate in Petri plates with 70%, 90% i 100% ethanol successively for 2 min in each concentration
- 13) Leave to dry at room temperature
- 14) Staining: place the slide into Petri dish and incubate in 5 mL of May-Grünwald dye for 6 min, wash with distilled water and transfer in next Petri dish, incubate in 10% Giemsa dye (4.5 mL deH<sub>2</sub>O + 0.5 mL Giemsa). Wash the dye under tap water and leave vertically to dry at room temperature
- 15) Examine the slides under microscope and count the spermatozoa
- 16) Classification:
  - spermatozoa without DNA fragmentation (big halo aura - equal to or bigger than  $1/3$  of the smaller core diameter, or medium halo aura – sized between the big and small halo aura)
  - spermatozoa with DNA fragmentation (small halo aura – similar to or smaller than  $1/3$  of the smaller core diameter, without halo aura, or without halo aura and degraded – irregularly and weakly stained core present, should not be counted)
- 17) Number of fragmented sperm is reported as % SDF ( $[\text{number of sperm with fragmented DNA} / \text{total sperm count}] \times 100$ )